

The Portuguese Beam Allocation Group at the ESRF: a 14-year old success history (1999-2013)



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2nd ENURS
15.02.2013

The ESRF at a glance

Founded in 1998

Began operation in 1994

Annual budget: ca. 80 M€

Members (minimum 4% shares, full voting rights):

France, Germany, Italy, United Kingdom, Spain, Switzerland, Benesync (Belgium & The Netherlands), Nordsync (Denmark, Finland, Norway, Sweden)

Scientific Associates (< 4% shares, limited voting rights):

*Austria, **Portugal**, Israel, Poland, Centralsync (Czech Republic, Hungary, Slovakia).*

Portugal, MX and the ESRF

The first Portuguese MX users of the ESRF were given access through a collaboration with the EMBL Grenoble Outstation in 1995-6

*Portugal joined the ESRF as a **scientific associate** in 1997 (1%).*

Novel statute created to accommodate the Portuguese membership, later allowed other small European countries to join the ESRF.

In 1999 the MX BAG scheme was created to promote a more efficient and productive use of the beamlines:

1 shift = 8 hours = MANY experiments

In recognition of their excellent work, the Portuguese MX groups were invited to form one of the first BAGs

Today, BAG use of the ESRF MX beamlines accounts for more than 90% of the available beamtime

The Portuguese MX BAG in 1999



ITQB – Universidade Nova de Lisboa

Maria Arménia Carrondo

REQUIMTE/FCT – Universidade Nova de Lisboa

Maria João Romão

IBMC – Universidade do Porto

Ana Margarida Damas

The Portuguese MX BAG in 2013



ITQB – Universidade Nova de Lisboa

Margarida Archer

Maria Arménia Carrondo

Carlos Frazão

Pedro Matias

IGC – Fundação Calouste Gulbenkian

Alekos Athanasiadis

REQUIMTE/FCT – Universidade Nova de Lisboa

Maria João Romão

IBMC/IMEB - Porto

Luís Gales Pinto

Sandra Macedo-Ribeiro

João Morais-Cabral

Pedro Pereira

The BAG scheme in Practice



*Beam allocation **every 6 months***

Yearly Report:** alternating **Progress Report** and **Full Report

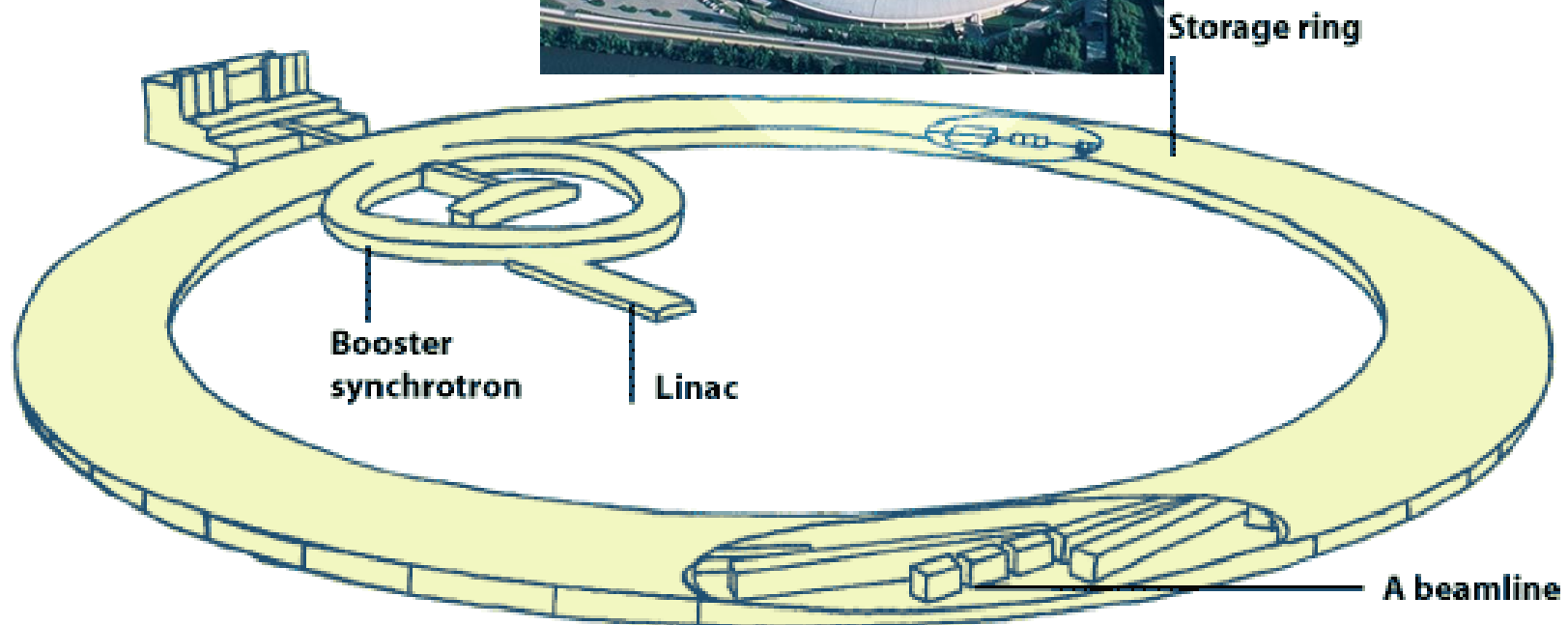
Report evaluation by Review Panel determines maintenance of BAG status and overall beam allocation

Our scores have been “good”;

To improve them to “excellent” in order to increase beam allocation, we need:

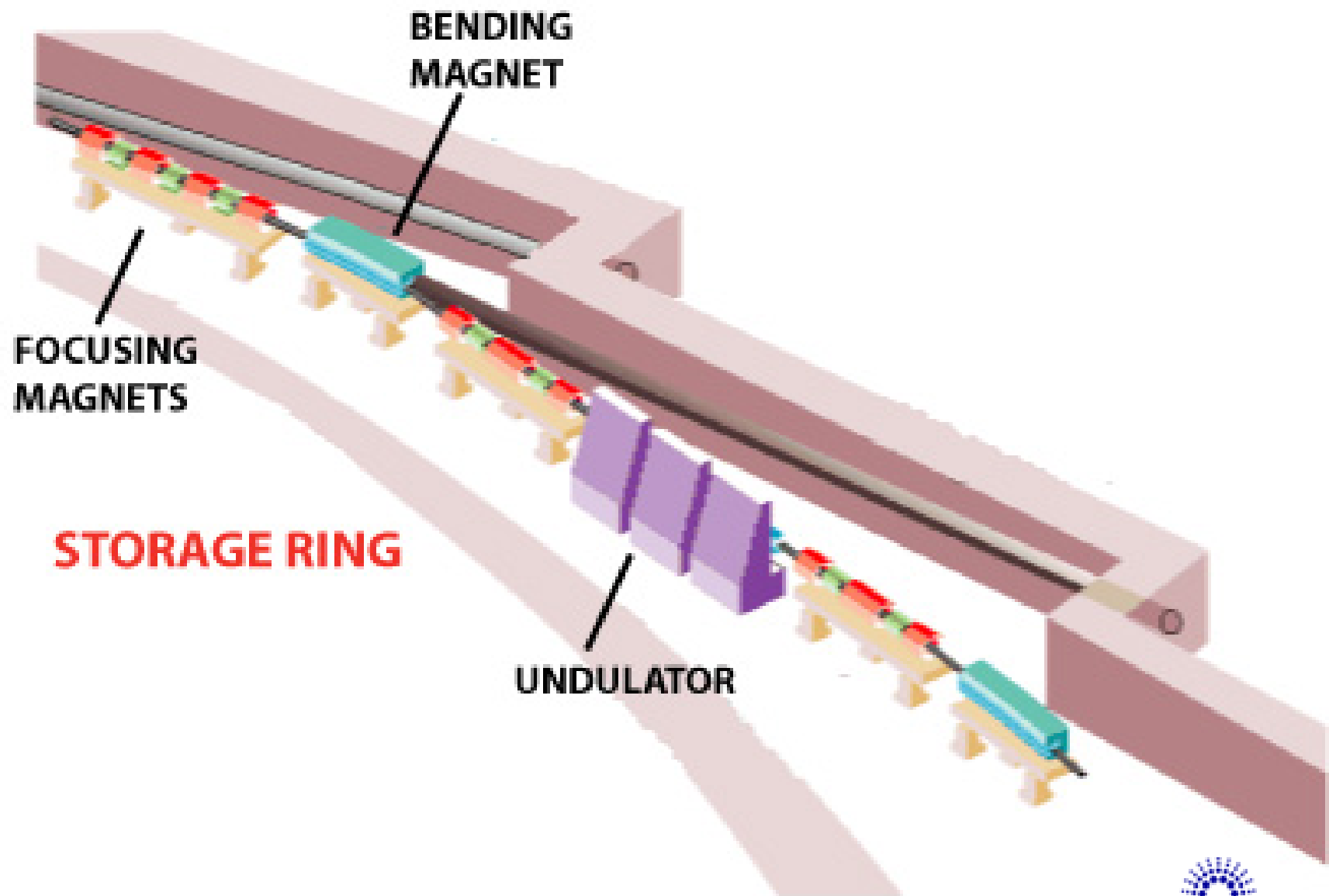
- *Work in more challenging projects (e.g., membrane proteins)*
- *More publications in high IF journals (e.g., Science, Nature, etc.)*

An Overview of the ESRF



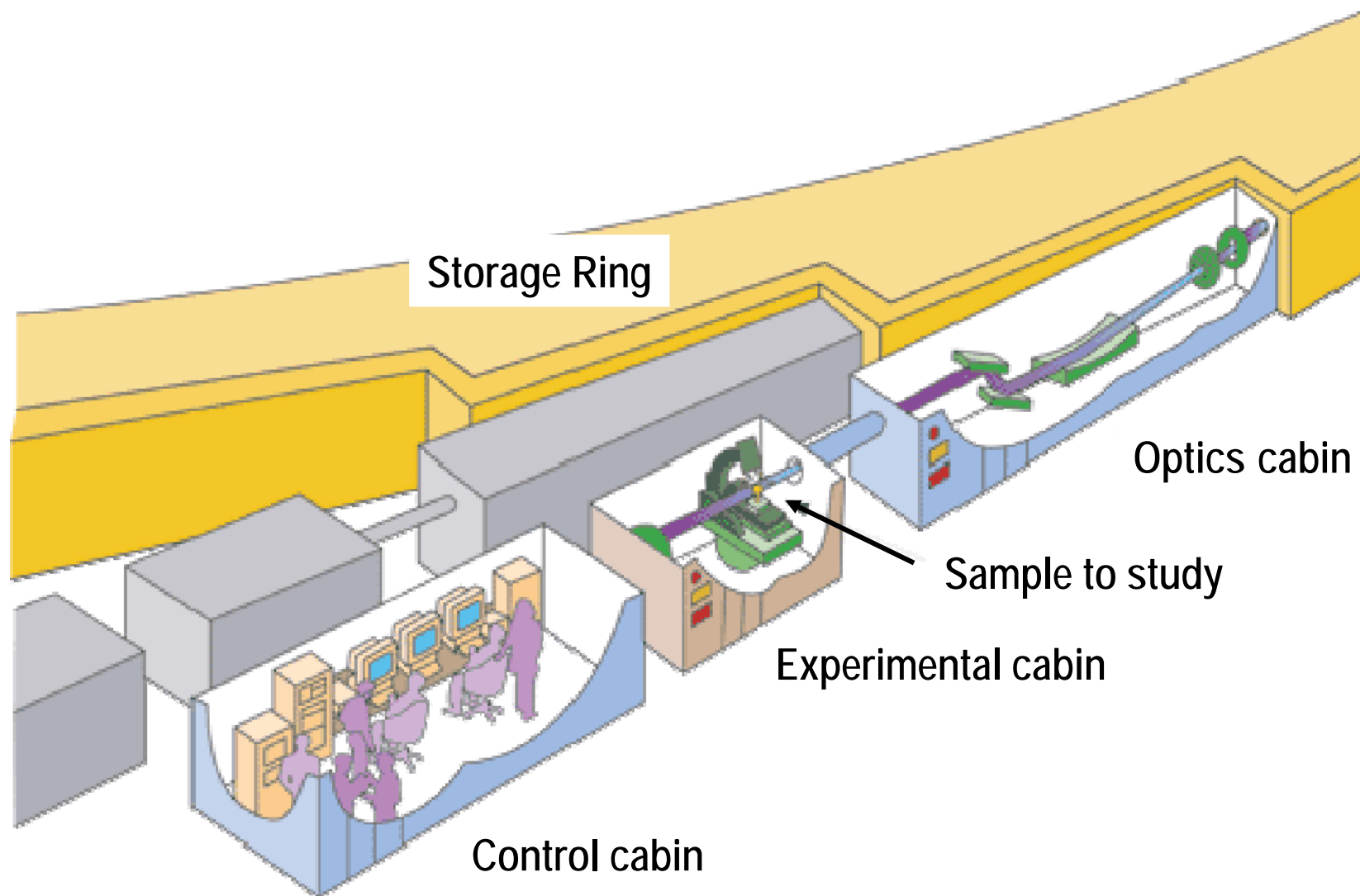
Source: <http://www.esrf.fr/AboutUs/GuidedTour/Animation>





Source: <http://www.esrf.fr/AboutUs/GuidedTour/Animation>

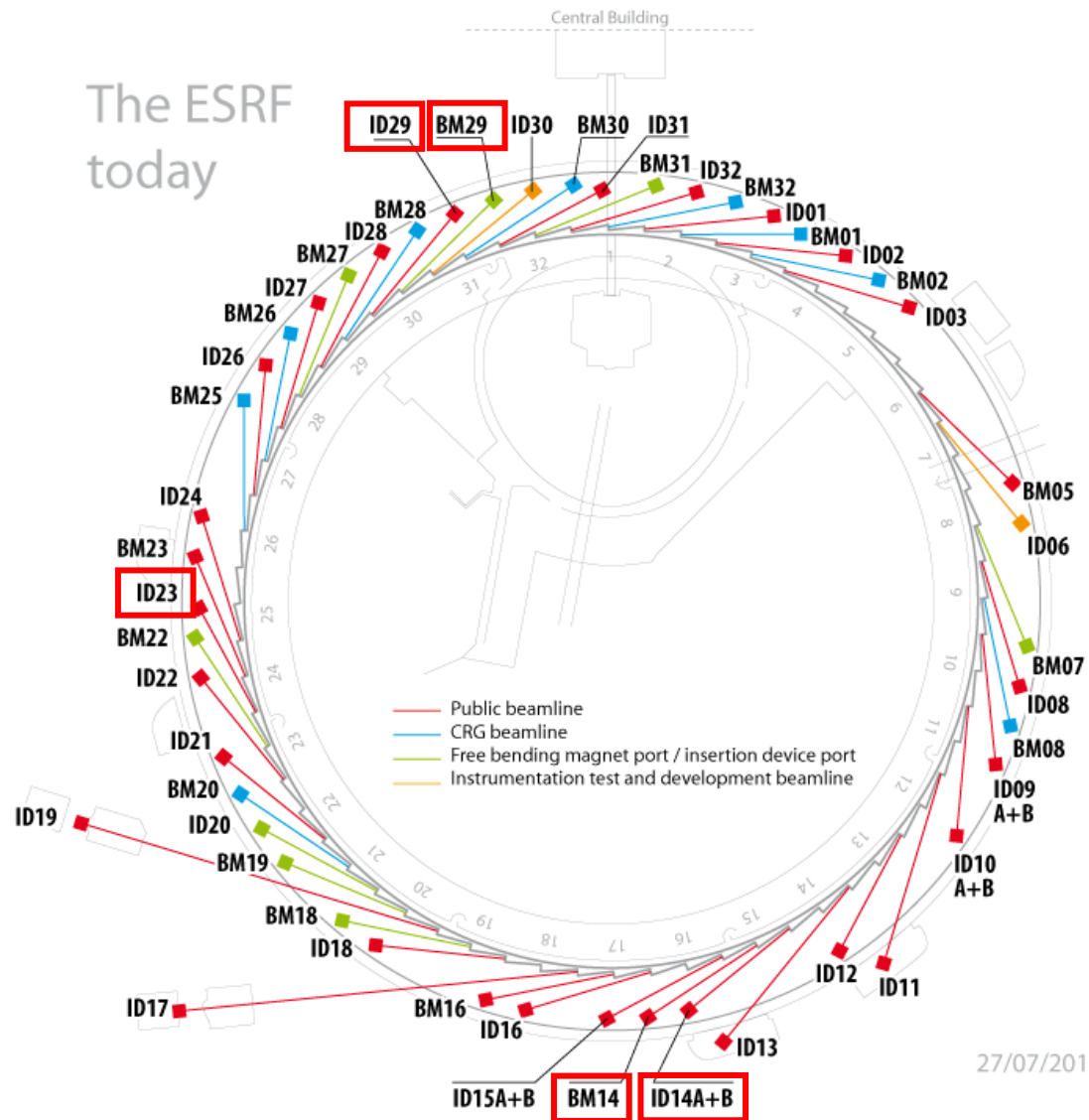




Source: <http://www.esrf.fr/AboutUs/GuidedTour/Animation>

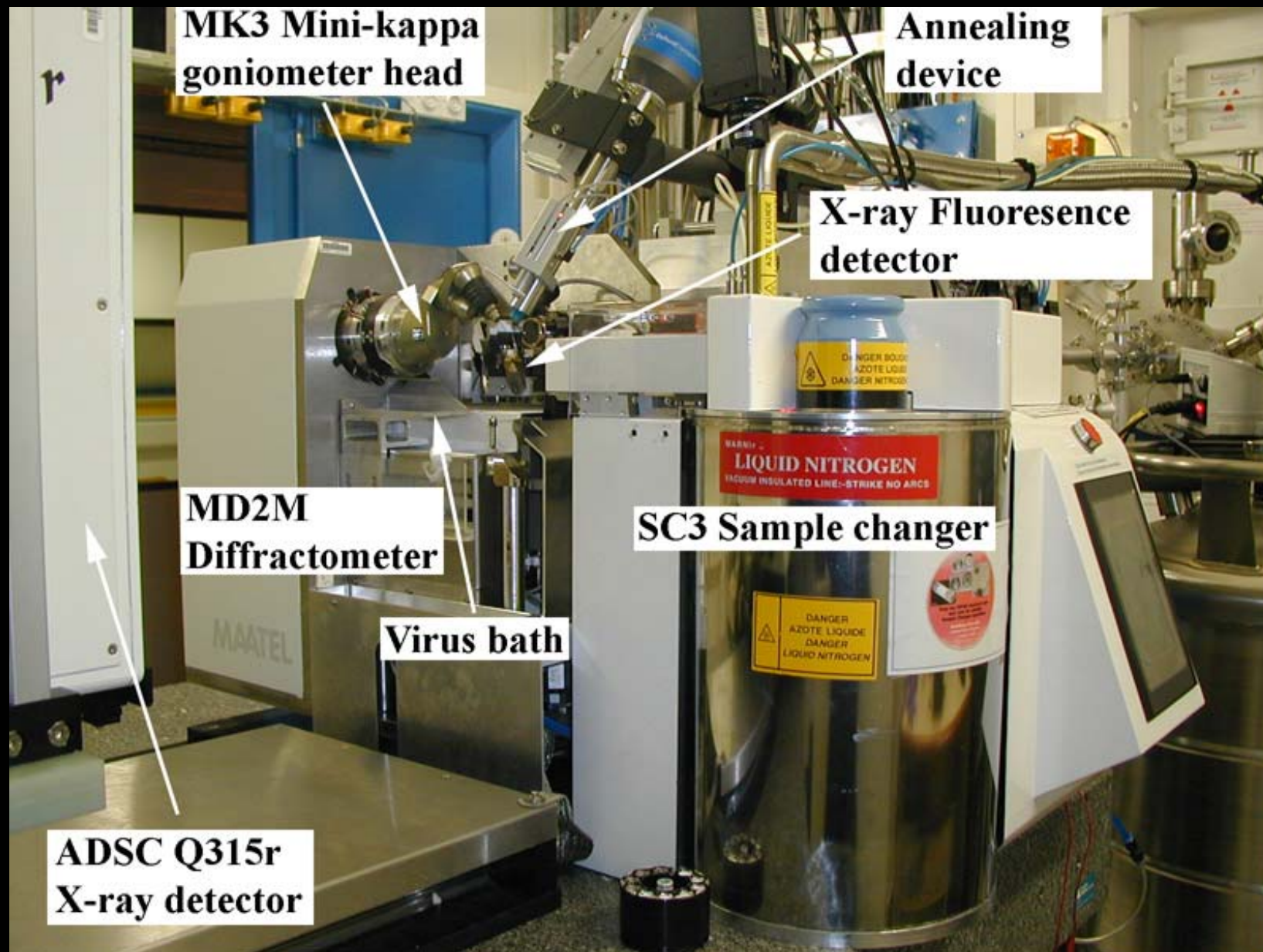


The MX beamlines at the ESRF



27/07/2011





**MK3 Mini-kappa
goniometer head**

**Annealing
device**

**X-ray Fluorescence
detector**

**MD2M
Diffractometer**

SC3 Sample changer

Virus bath

**ADSC Q315r
X-ray detector**

mxCuBE (opid-291) Expert mode

File Instrumentation Help

Match Collect Energy scan XRF spectrum Image EDNA Log

Motor height: 275.4 21.94 1.0

Available motors: Phi Z 0.034 1.0

Minkappa: omega 275.4 kappa 0.000 phi 24.010
 -X: 0.232 Y: 0.528 Z: 0.041

Commands: Initialize Home Manual Load

Energy: Current: 22.247 kvv 0.573 A Transmission: 99.110
 Move to: keV Set to: Filters

Beamstop: out Safety shutter: closed Fast shutter: closed

Scan plot: 01. 628.794946, Y. 1202.970207

Grid: Nb points: points per line steps (mm) distance
 Axis 1: 10 0.04 0.41
 Axis 2: 15 0.02 0.26

Motor 1: phi y Motor 2: phi z Move to start Fill queue Stop

Information messages: submit feedback chat spec EDNA log

2010-06-10 13:24:21 Got the following error from the ISPyB server: No samples whatsoever for proposal #7, session 27410, bar codes []
 2010-06-10 13:24:21 Got the following error from the ISPyB server: No samples whatsoever for proposal #7, session 27410, bar codes []
 2010-06-10 13:24:47 scale 539.665407 544.365814
 Jun 10 09:07 Delivery: Next Refill at 21:00

Sample centring: User confirms Centre Accept Reject Autofocus Snapshot

Sample display: Light: 22.0 1.0 Front Light: 28.0 5.0 Zoom: 5 Focus: -0.01 0.01

Sample centring has finished. - Grid grab, select the area

Beam size: Hsv 0.5 Vsv 0.2 Move Off-sets Aperture: Diameter: 75

Machine current: 179.7 mA 78 mA/turn/h

Cryo: 100.0 K

Dry: unknown
 Supervisory: unknown
 Logging: unknown

Culter / Stage: 1. Preparing beamline
 2. Mounting sample
 3. Centring sample
 4. Centring images

ADSC temperature: cooling ok

Photon flux with 50 um Aperture flux: 0 phys

Current users: Selecting gives control Allow timeout control Take control My name: jrl-479

spec.exe

Taskbar: mxCuBE Front End & Vacuum ISPyB BioSync | Statistics by Year Calculator Shell - Konsole <3> idappli ID Control Vacuum Viewer 1.10 [users: mxCuBE (opid-291) Shell - Konsole <2>

System tray: 13:28 10/06/2010

NX - leonard@tet.esrf.fr:1137 - remote

File New Bookmarks Desktop Windows Help

File Information Help

Home Collect Home of mine File spectrum Image dna

User: **mx-415 TEST** (MCCARTHY.E.S.R.F. Date: 2008-04-20 to 2008-05-02)

Available samples:

Name	Acronym	Barcode	Location	Space group	a	b	c	α	β	γ	Minutes	Basket
		HADDA01400		300								
		HADDA00751		302								
		HADDA17605		306								
		HADDA07207		305								
		HADDA00736		300								
		HADDA07207		307								
		HADDA17615		306								
		HADDA17620		309								
		HADDA07200		333								
	FAE	CADDA01001		300	P212121	65.4	208.8	113.9	90.0	90.0	0.0	0.0

Show only the samples inside the sample changer (30 samples, 51 baskets)

Group by: No grouping Refresh

Energy: 11.288 keV Wavelength: 0.109 Å

Current: 0.0 mA Set to: 0.0 mA

Parameters: Queue (0) Status

Directory: /data/2008/04/20/20080420

Profile: mx415 Oscillation start (deg): Oscillation range (deg): Overlap (deg): Exposure time (s): Number of images: 1 Number of passes: Comments:

Collect data Add to queue

Sample changer: Standby

Ready to open

Current basket: Position: 5 Scan

Current sample (HADDA02205): Samples in queue: 4 Position: 4 Scan

Upfront HADDA02207

Basket 1: 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30

Basket 2: 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30

Basket 3: 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30

Basket 4: 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30

Basket 5: 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30

Double click loads the sample

Scan selected baskets

Machine current: 0.0 mA

Collect stage:

1. Preparing beamline
2. Mounting sample
3. Centering sample
4. Collecting images

Current users: jmx415@b2

Selecting gives control

Allow timeout control

My name: jmx415-2

100:24:14 I've joined the conversation as jmx415-2 (operator in jmx415@b2).

100:24:14 jmx415@b2 has control.

100:24:14 jmx415-2 wants to have control.

100:24:15 jmx415@b2@jmx415-2 will have control in 30 seconds...

100:25:00 jmx415@b2@jmx415-2 will have control in 20 seconds...

100:25:10 jmx415@b2@jmx415-2 will have control in 10 seconds...

100:25:24 I've gained control.

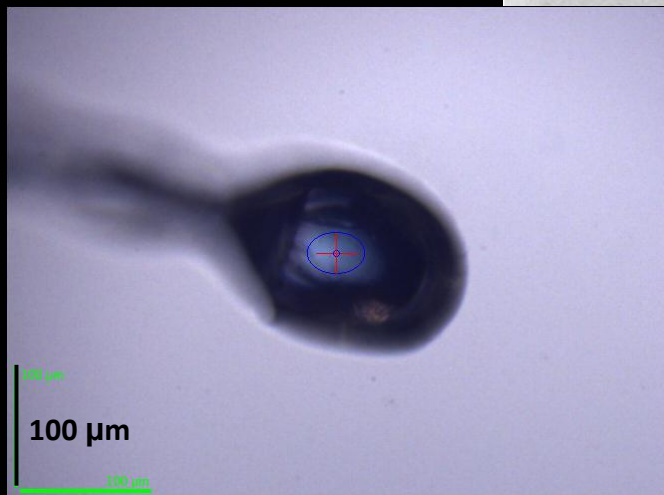
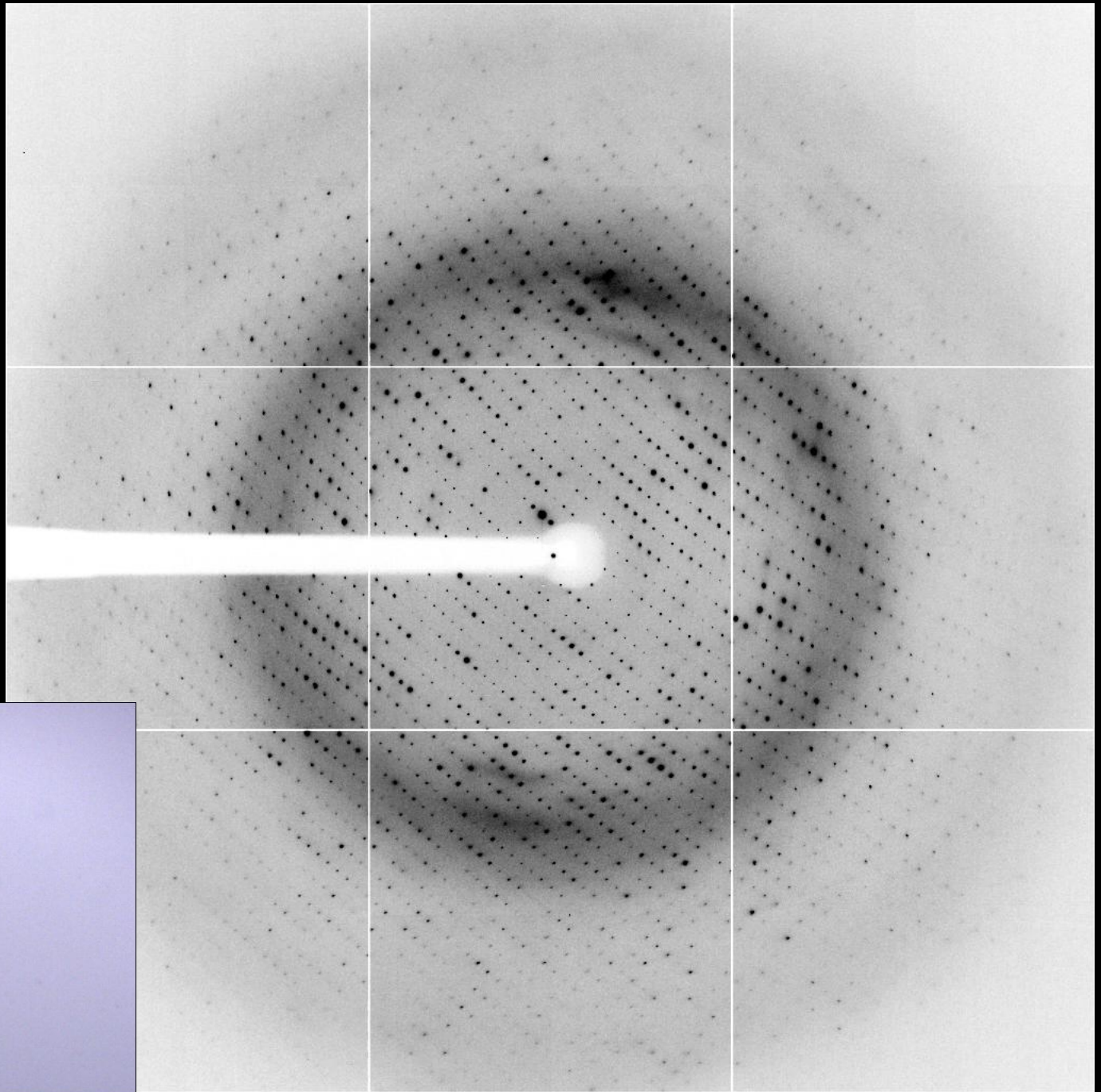
Say:

Apr 30 07:44 51° cavity conditioning in progress. No beam expected before 10:00.

Shell - Konsole

mxCuBE (mx-415)

Start Eudora NX - leonard@tet.esrf... tfranView 09:27



Firefox | MXcube - Pesquisa do Google | Resultados da pesquisa de htt... | View results

https://www.esrf.fr/ispyb/user/viewResults.do?reqCode=display&dataCollectionId=1070465&rmerge=10.0&isigma=1.0&anomalous=false

Mais visitados | Anomalous Scattering C... | Best of the Web | Channel Guide | Customize Links | Free Hotmail | Internet Start | Is Your Operating System... | Microsoft | MSN.com | Windows Marketplace

View last sessions
View all sessions
Search data collections

Selected Session		Selected DataCollectionGroup		Selected DataCollection	
Start Date	26-01-2013	Start Time	29-01-2013 06:02:00	Start Time	29-01-2013 06:02:00
BeamLine	ID14-4 +33470882322	Experiment Type	SAD	Image Prefix	ref-A32
	Back to this session		Back to this data collection group	Run Number	1
	Back to sessions				Back to this data collection

Experiment parameters | Beamline parameters | Characterisation parameters | **Characterisation results** | Autoprocessing

Characterisation Results ([EDNA log file](#))

Data collection info

Data collection date	Tue Jan 29 06:02:34 2013
Image prefix	ref-A32_1
Directory	/data/visitor/mx1428/id14eh4/20130127/RAW_DATA/IBMC/AMPPNP06b

Diffraction Plan

Forced space group	Anomalous data	Aimed multiplicity	Aimed completeness	Aimed I/sigma at highest res.	Aimed resolution (Å)
None	False	Default (optimized)	Default (>= 0.99)	2.00	Default (highest possible)

Collection plan strategy ([RADDPOSE log file](#) , [BEST log file](#))

Resolution limit is set according to the given max.time

Wedge	Subwedge	Start (°)	Width (°)	No images	Exp time (s)	Max res (Å)	Rel trans (%)	Distance (mm)
1	1	131.00	0.20	705	0.10	1.87	67.40	267.64

Strategies Wedge

Wedge number	Resolution Å	Completeness	Multiplicity	Total dose	Number of images	Phi	Kappa	Wavelength	Comments
1	1.87	99.3	3.0		705			1.0	

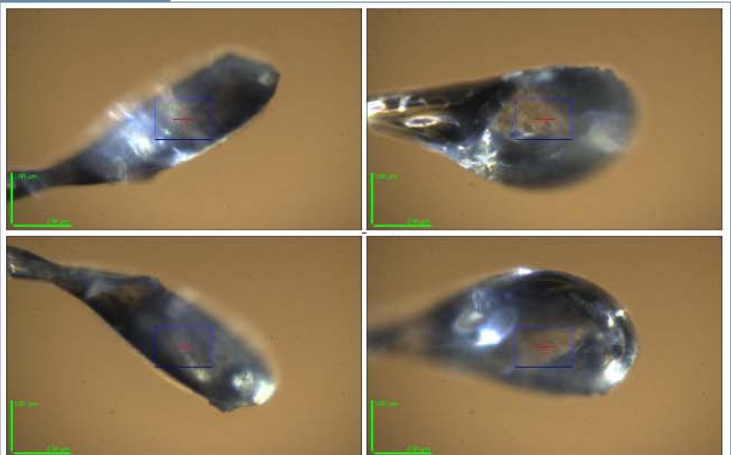
Windows taskbar: 14:03 11-02-2013

Strategies Wedge

Wedge number	Resolution A	Completeness	Multiplicity	Total dose	Number of images	Phi	Kappa	Wavelength	Comments
1	1.87	99.3	3.0		705			1.0	

Sub Wedge number	Rotation axis	Axis start	Axis end	Exposure time s	Transmission %	Oscillation range	Completeness	Multiplicity	Total dose	Number of images	Comments
1		131.0	272.0	0.1	67.4	0.2				705	

Crystal Snapshots



Expected Snapshots location: /data/pyarch/id14eh4/mx1428/20130127/RAW_DATA/IBMC/AMPPNP06b/ref-A32_1_1.snapshot.jpeg

Images collected





- View last sessions
- View all sessions
- Search data collections

Selected Session

Start Date: 20-01-2013
 BeamLine: ID14-4 +33476882322
[Back to this session](#)
[Back to sessions](#)

This page is still under development and will gradually be updated.

Session info...

Session Informations

Local contact:

Session Comments:

[View dataCollections for all groups](#)

DataCollection Group	Protein Acronym (Image prefix)	Sample name	Sample position	Start Time	Comments	Nb Data Collections (Nb images)	View collections
SAD	(A32w1)		7 (3)	29-01-2013 06:04:25	Resolution limit is set according to the given max.time	1 (705)	
SAD	(ref-A32)		7 (3)	29-01-2013 06:02:00	Image created for EDNA characterisation	1 (4)	
SAD	(ref-A36)		8 (3) HA00AN4734	29-01-2013 05:57:09	Image created for EDNA characterisation	1 (4)	
SAD			4 (3)		Image created for EDNA characterisation	1	



- View last sessions
- View all sessions
- Search data collections

Selected Session	Selected DataCollectionGroup
Start Date: 29-01-2013 BeamLine: ID14-4 +3347882322 Back to this session Back to sessions	Start Time: 29-01-2013 06:04:25 Experiment Type: SAD Back to this data collection group

Data Collection info...

Reports	Parameters
View DOC report View PDF report View CSV report DOC Screenings PDF Screenings	Ignore RSymm in the low resolution shell over: <input type="text" value="10.0"/> Ignore I / Sigma in the low resolution shell under: <input type="text" value="1.0"/> <input type="button" value="Update"/>

Image Prefix	Run No	Protein Acronym	Start Time	# images	Experiment Parameters (Expand)	Status	Space Group	Completeness	Resolution	Rsymm Inner Outer Overall	Unit cell a b c alpha beta gamma	Sample Ranking	Skip	Comments
		All										Rank <input checked="" type="checkbox"/> EDNA	<input type="checkbox"/>	Save
A32w1	1		29-01-2013 06:04:26	705		●●●●	P 1 21 1	<div style="width: 100%; height: 10px; background-color: green;"></div>	100.0 - 4.22 2.03 - 1.96 100.0 - 1.96	2.8 62.0 4.6	36.8, 59.3, 71.9 90.0, 99.5, 90.0	<input type="checkbox"/>	<input type="checkbox"/>	
		All										Rank	<input type="checkbox"/>	Save



- View last sessions
- View all sessions
- Search data collections

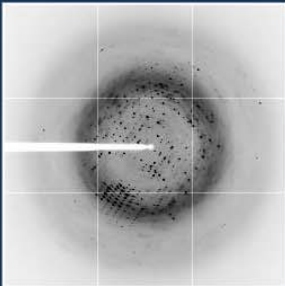
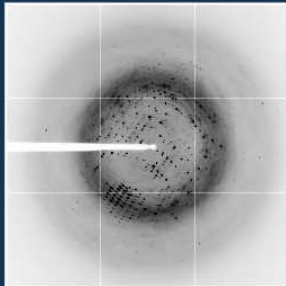
Selected Session	Selected DataCollectionGroup	Selected DataCollection
Start Date: 20-01-2013 BeamLine: ID14-4 +33476882322 Back to this session Back to sessions	Start Time: 29-01-2013 06:04:25 Experiment Type: SAD Back to this data collection group	Start Time: 29-01-2013 06:04:26 Image Prefix: A32w1 Run Number: 1 Back to this data collection

Experiment parameters	Beamline parameters	Autoprocessing
Synchrotron name ESRF		
Synchrotron filling mode 7/8 multibunch		
Beamline name ID14-4		
Undulator types u35 u23 u24		
Undulator gaps 15.71 mm 16.01 mm 17.18 mm		
Beam transmission 74 %		
Slit gap Hor 100 µm		
Slit gap Vert 100 µm		
Detector type CCD		
Detector name q315r		
Detector manufacturer ADSC		
Detector mode HARDWARE BINNED		
Detector pixel size Hor 0.1026 mm		
Detector pixel size Vert 0.1026 mm		
Focusing optics Toroidal mirror		
Monochromator type Si(111)		
Beam shape rectangular		
Flux 2.51E12 photons/sec		
Flux end 2.47E12 photons/sec		
Beam size at sample Hor 100 µm		
Beam size at sample Vert 80 µm		
Beam divergence Hor 12 µrad		

Crystal Snapshots

Expected Snapshots location:

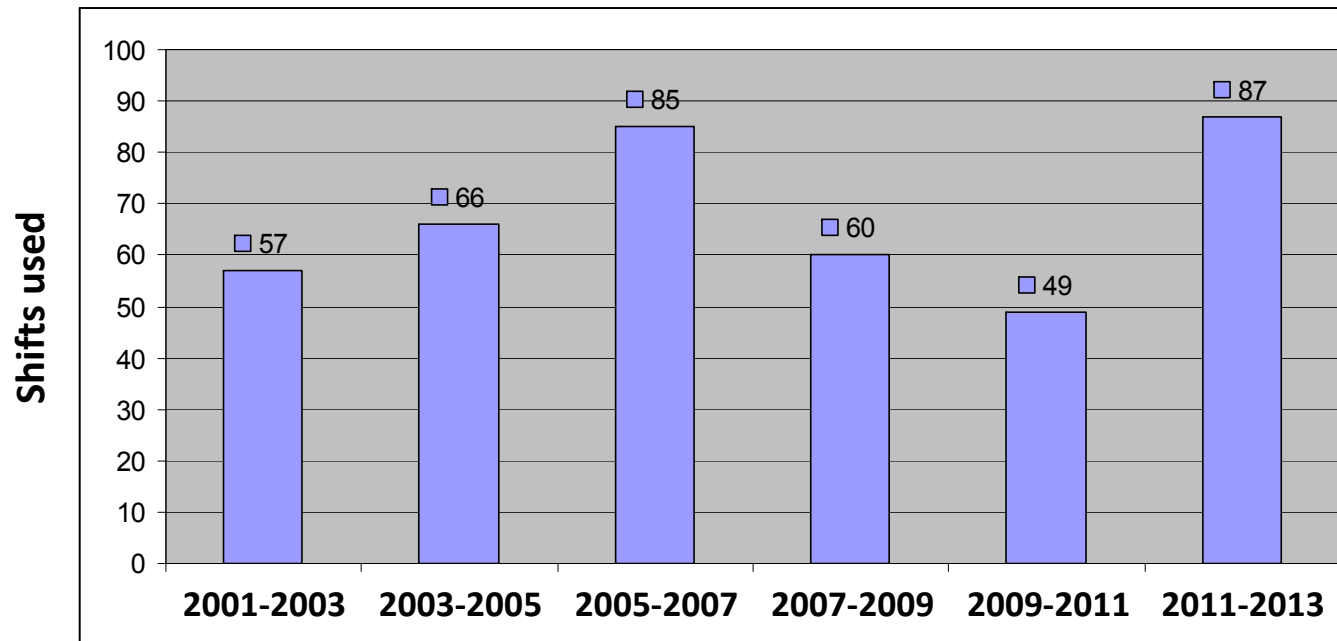
Images collected

	
Machine message: Jan 28 21:05 No scheduled refill until tomorrow 8h00. (M.D.T.)	Machine message: Jan 28 21:05 No scheduled refill until tomorrow 8h00. (M.D.T.)
Comments:	Comments:

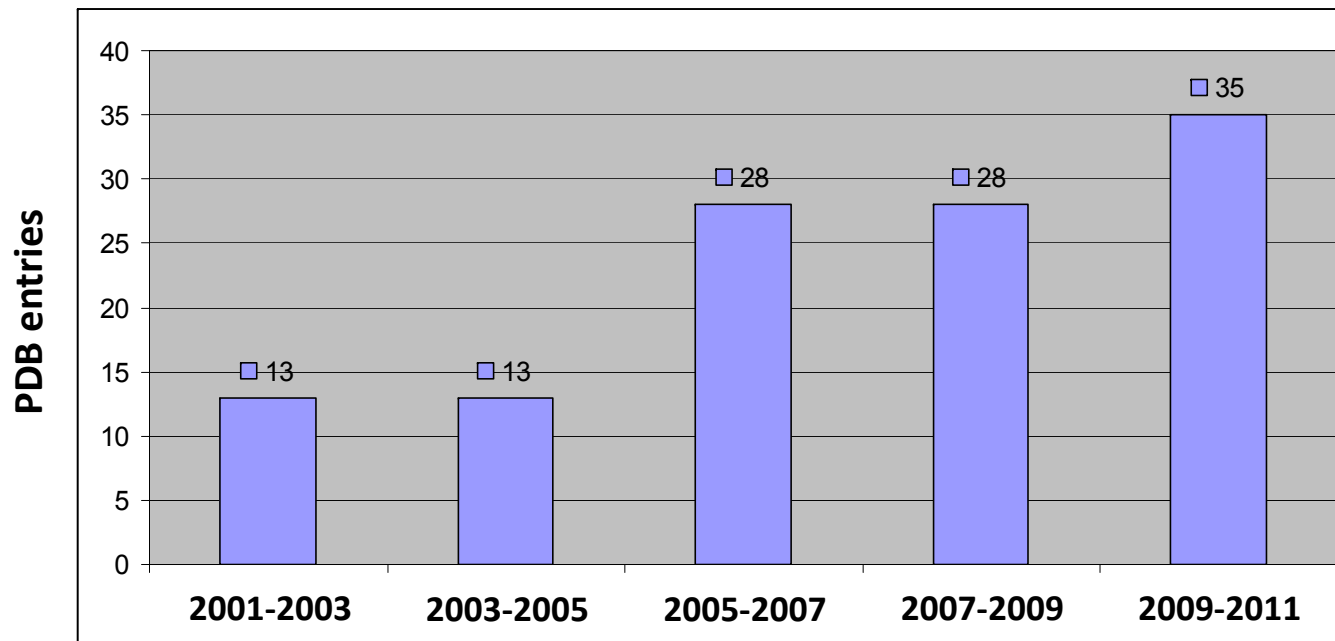
Location: /data/visitor/mx1428/id14eh4/20130127/RAW_DATA/IBMC/AMPNP06b

[View as Image wall](#)

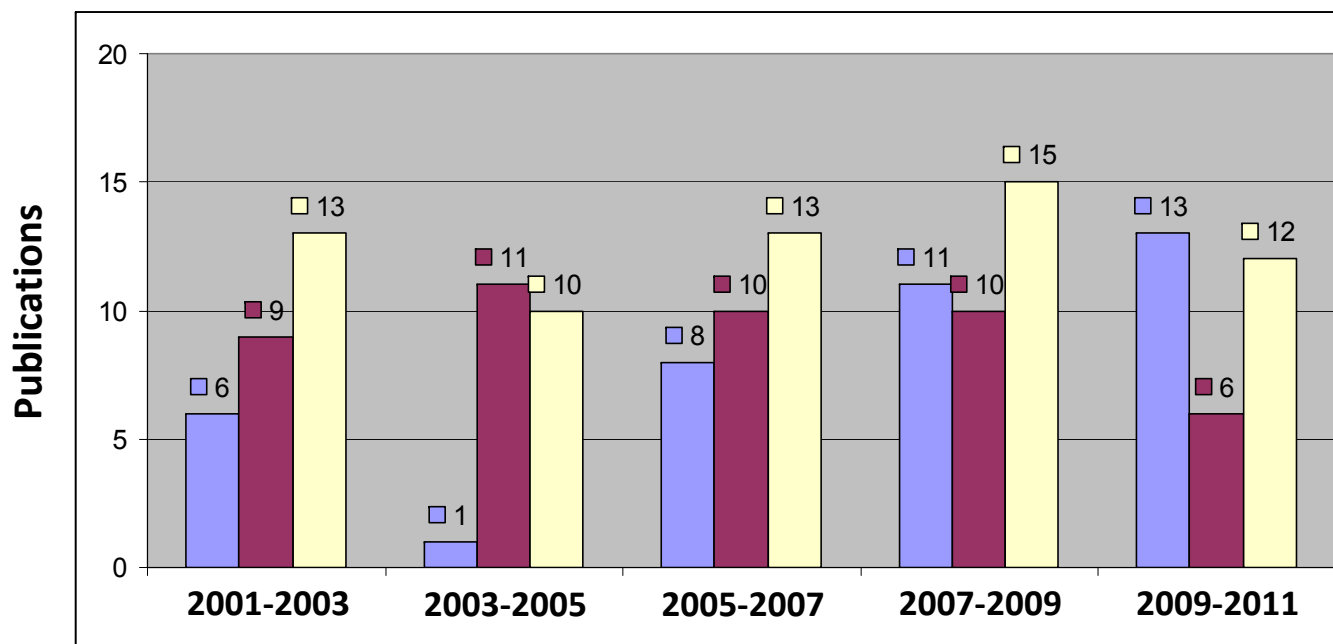
BAG statistics - shifts used



BAG statistics - PDB depositions



BAG statistics - Publications

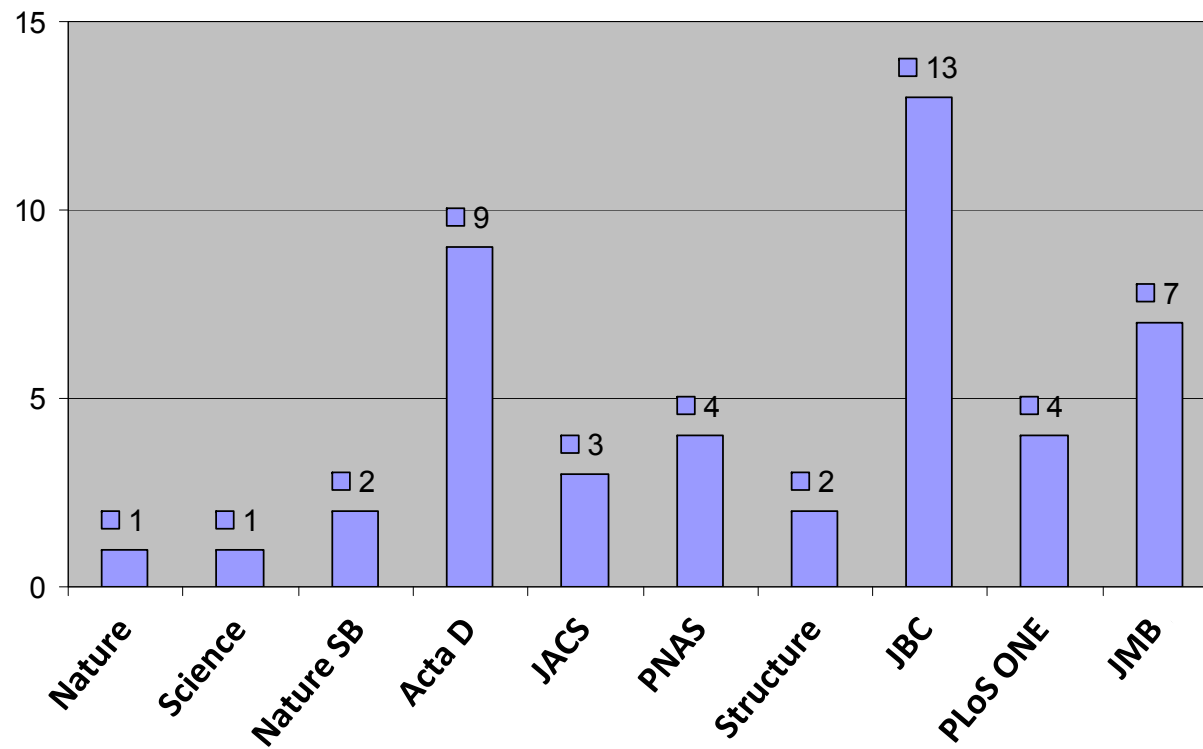


■ Crystallization Reports

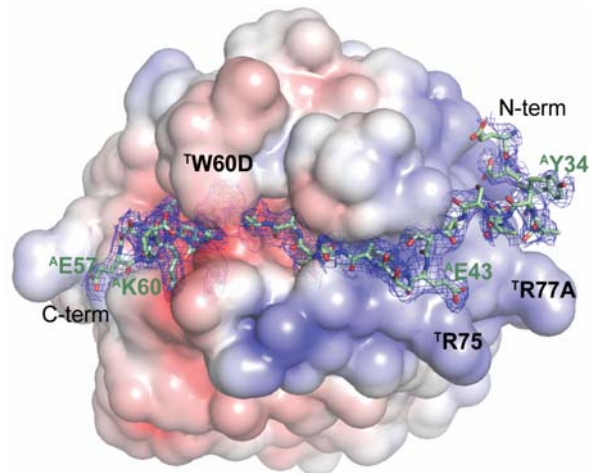
■ Journals with IF > 4

■ Journals with IF < 4

Publications with IF > 4: 2001-2011



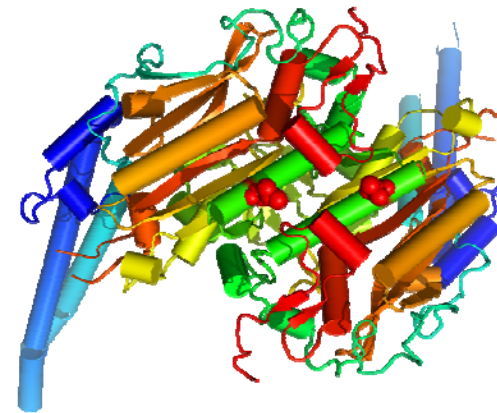
Novel anticoagulant mechanism in the malaria mosquito



- Anophelin binds to thrombin in the reverse direction of *bona fide* substrates disrupting the catalytic triad
- Its compact size and resistance to proteolysis might the design of novel antithrombotics

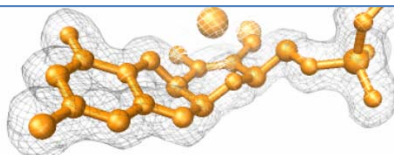
Figueiredo *et al.* (2012) *Proc. Natl. Acad. Sci. USA* **109**, E3649-58

SerRS, the main player in *C. albicans* genetic code alteration



- Crystal structures of the two natural isoforms of *Candida albicans* seryl-tRNA synthetase (SerRS-Leu / SerRS-Ser)
- Ambiguous codon localization tailored to minimize protein misfolding events

Rocha *et al.* (2011) *Proc. Natl. Acad. Sci. USA* **108**, 14091-6



Main Research lines & Highlights

1- Molybdopterin Enzymes

- Aldehyde Oxidases:

The first **mammalian aldehyde oxidase** structures (mouse and human)

- Detailed **mechanistic studies and novel mechanisms** based on atomic-resolution structures (MOP, NAP)

2- Drug Design

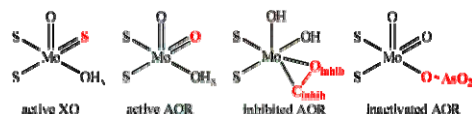
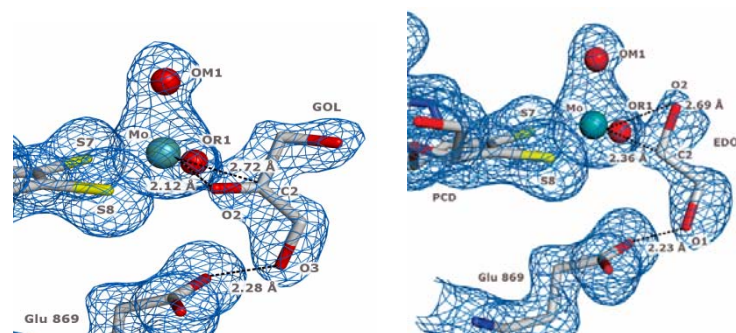
- CO Releasing Molecules (CORM) binding mechanism to plasma proteins
- Adducts of Human Transferrin and Vanadium as anti-diabetic agents
- Design of potent and specific inhibitors of proteases based on Trypsin studies

3- Cellulosome: a megaDalton complex for cellulose degradation:

- Carbohydrate binding Modules
- Glycoside hydrolases
- Cohesin-dockerin complexes

- Detailed mechanistic studies
- Ligand identification and novel mechanisms

Ethylene-glycol & Glycerol inhibited AOR @1.7 Å

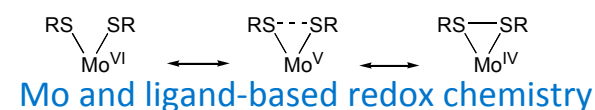
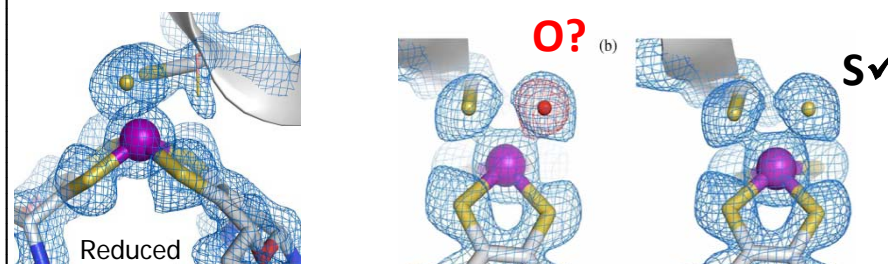


- First structural evidence for a Mo-C bond in a biological system
- Possible to distinguish Mo-O and Mo-C bonds
- Substrates may also bind to the protein by a η^2 coordination

Romão, *Dalton Trans*, 2009
Santos-Silva et al, *JACS*, 2009
Correia et al, submitted

POSTER & Oral communication

Oxidized & Reduced Nitrate Reductase @1.5 Å



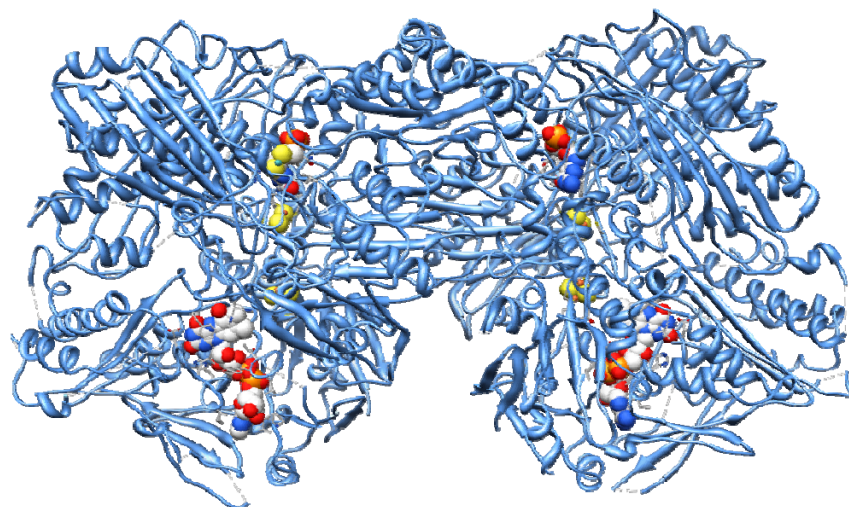
- New mechanism for nitrate reduction:
Ligand- based redox chemistry
- Unanticipated sulfur ligand
- Partial disulfide bond

Coelho et al, *J Mol Biol*, 2011

- First Crystal Structures of mammalian Aldehyde Oxidases
- Identification of new metallic clusters

Mouse Aldehyde Oxidase 3.0 Å

@ ID14-1 & ID23-1



Extremely poor crystals and weak diffraction!

(P1 (4 mols/au; 1300 aa))

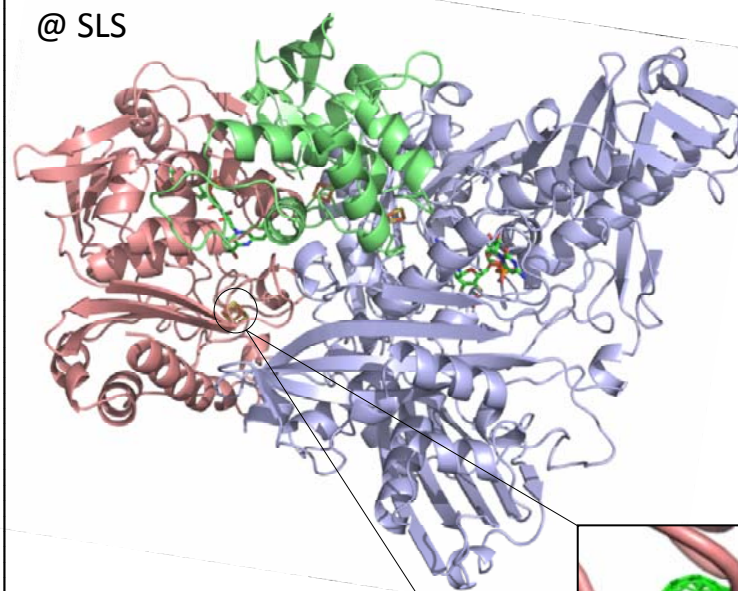
- Insights into substrate specificity
- Important in drug metabolism & increasing importance in recent drug design programs (Pfizer)

Coelho et al, *Drug Metab Dispos.* 2011

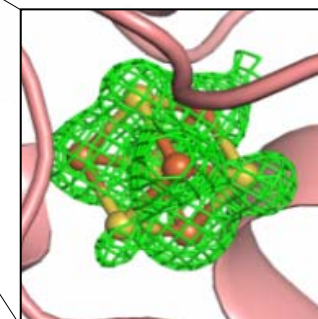
Coelho et al, *J Biol. Chem.* 2012

E. coli Periplasmic Aldehyde Oxidase 1.8 Å

@ SLS



Identified a new [4Fe-4S] cluster!



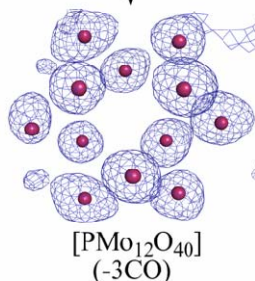
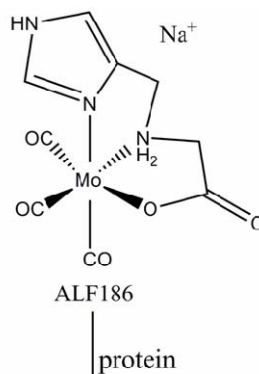
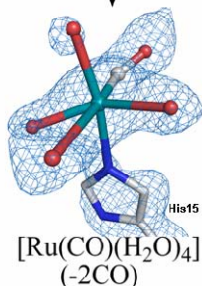
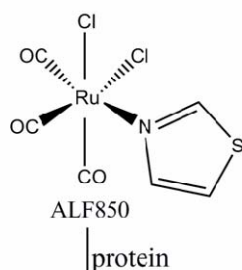
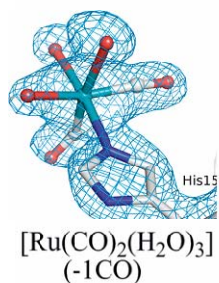
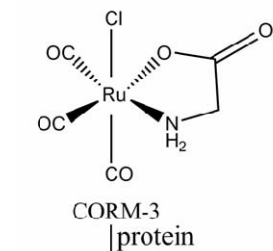
- The first monomeric XO-related enzyme
- Unique member of the XO family

POSTER

- CO Releasing Molecules (CORM) binding mechanism to plasma proteins
- Vanadium – Transferrin adducts
- Trypsin – Inhibitors binding

CORM – Lysozyme adducts 1.7 Å

@ ID14-1 & ID23-1



- Structural characterization of protein – CORM interactions
- Insights into CO release and polyoxometallate formation

Santos-Silva et al, *JACS*, 2011; *Curr Med Chem*, 2011

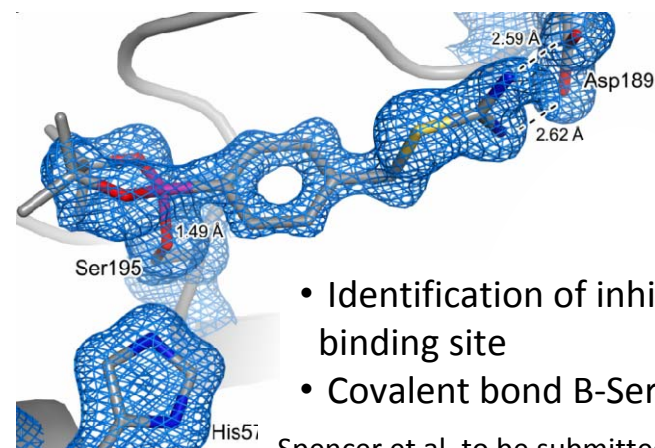
Santos et al, *JIB*, 2012

Seixas et al, *Dalton T*, 2012

POSTER

Trypsin and inhibitors of Urokinase 1.5 Å

@ ID29



- Identification of inhibitors binding site
- Covalent bond B-Ser195

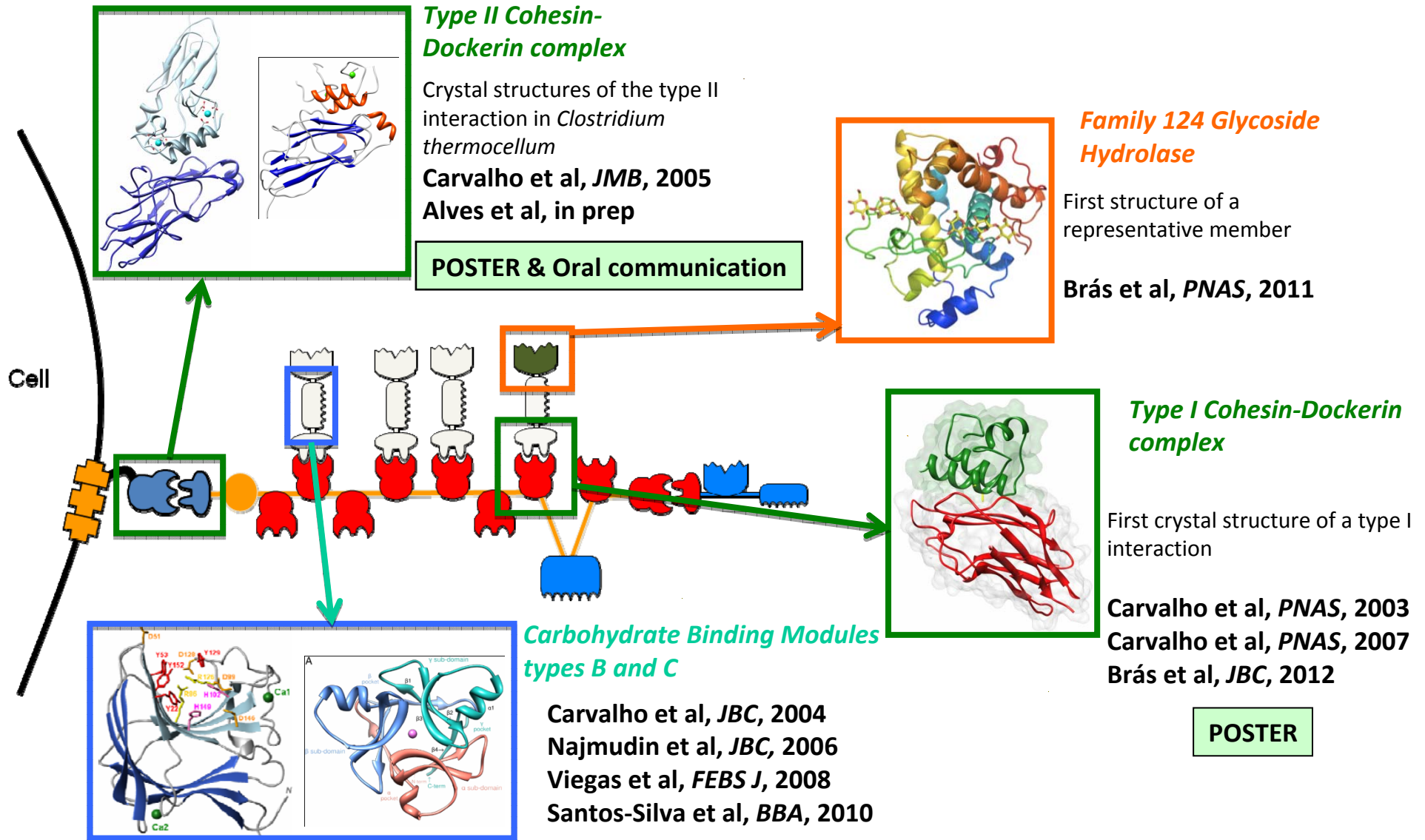
Spencer et al, to be submitted

Vanadium - Human Transferrin adducts

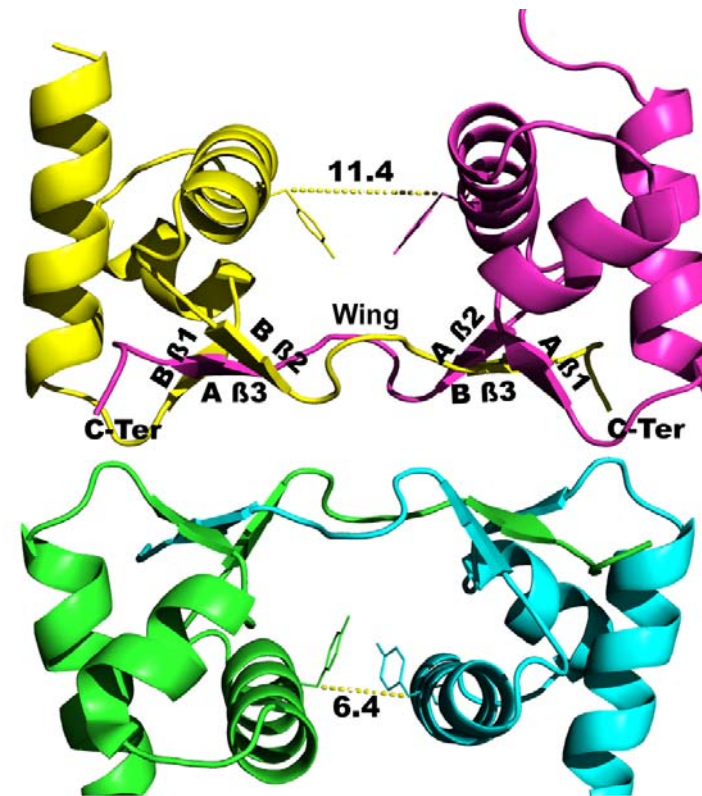
- Identification of the binding sites of V compounds
- Conformational changes upon binding (SAXS)
- Urea Gel Electrophoresis to control protein modifications upon binding

Mehtab et al, *J Inorg Biochem*, in press

Cellulosome: a megaDalton complex for cellulose degradation



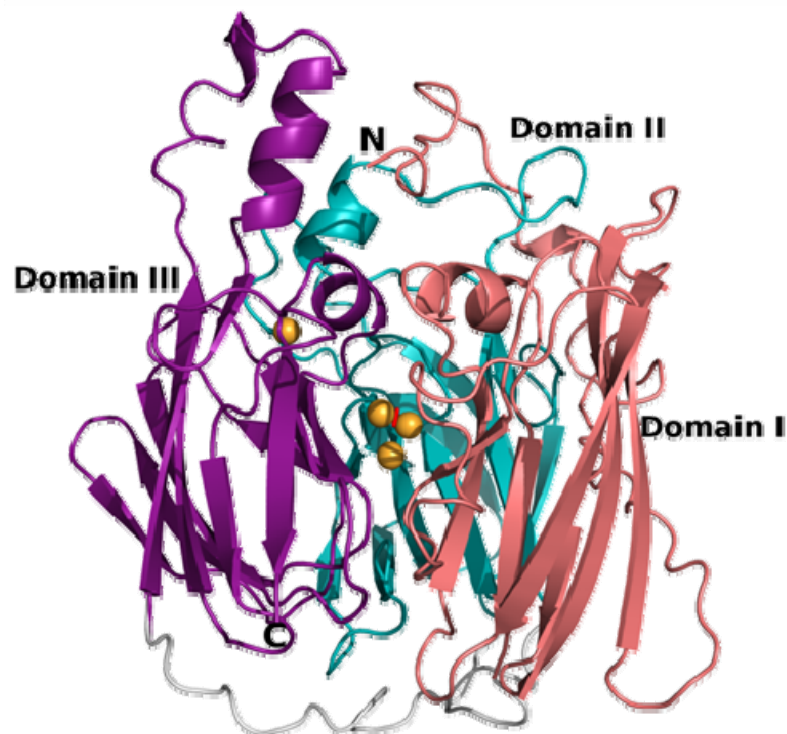
Cyprinid Herpes Virus 3 Orf122 Crystal Structure - A poxvirus-like Zalpha domain



Unlike other Zalpha domains, ORF112 forms a dimer through a unique domain-swapping mechanism.

Tomé AR, Kuš K., Correia S., Paulo L., Zacarias S., de Rosa M., Figueiredo D., Parkhouse RME. and Athanasiadis A. (2013) J. Virology in print

Structure Characterization of a multicopper oxidase from *Campylobacter jejuni* CGUG11284



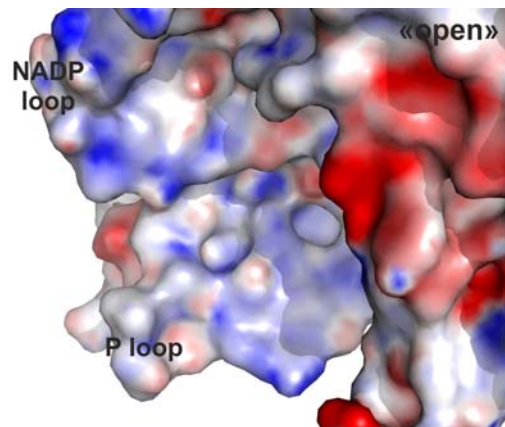
Resolution 1.95 Å

Campylobacter jejuni is a Pathogenic, Gram-negative bacterium, that is the most common cause of human gastroenteritis and bacterial food poisoning.

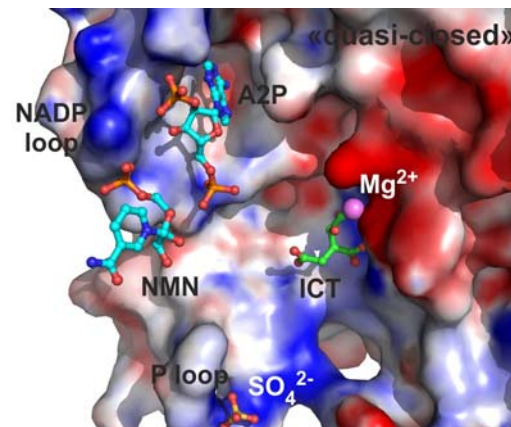
McoC – is a periplasmic multicopper oxidase thought to be involved in copper homeostasis.

The McoC structure displays a characteristic laccase-like fold, with three cupredoxin domains and two types of copper centres: a **T1** copper centre in domain III and a tri-nuclear center, with two **T3** and one **T2** copper atoms, localised between domain I and III.

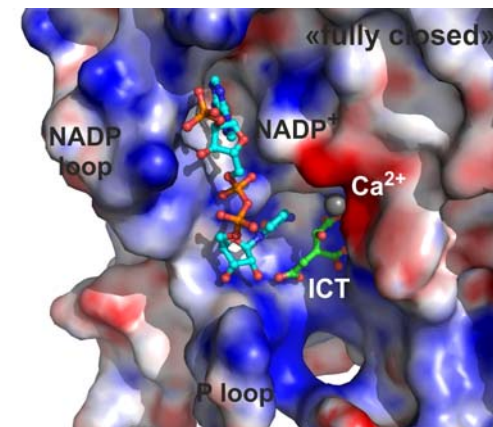
Induced Fit and the Catalytic Mechanism of Isocitrate Dehydrogenase



PDB 1sjs
Finer-Moore et al. (1997)



PDB 4ajs, ESRF ID23-1
Gonçalves et al. (2012)



PDB 4aj3, *in-house*
Gonçalves et al. (2012)

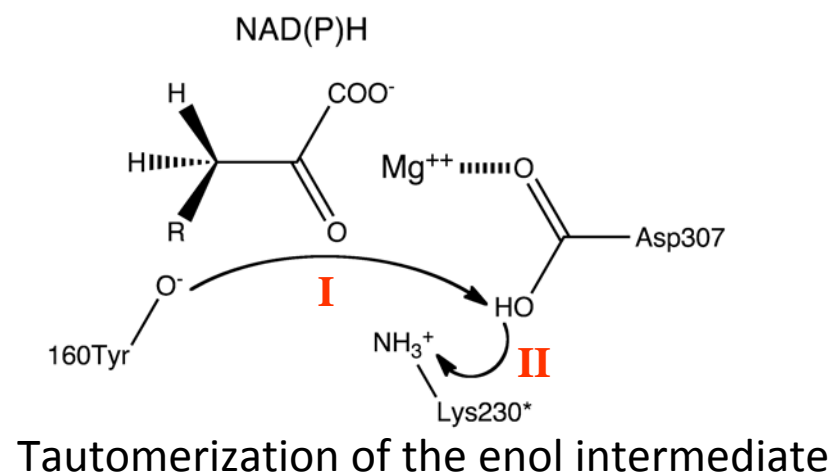
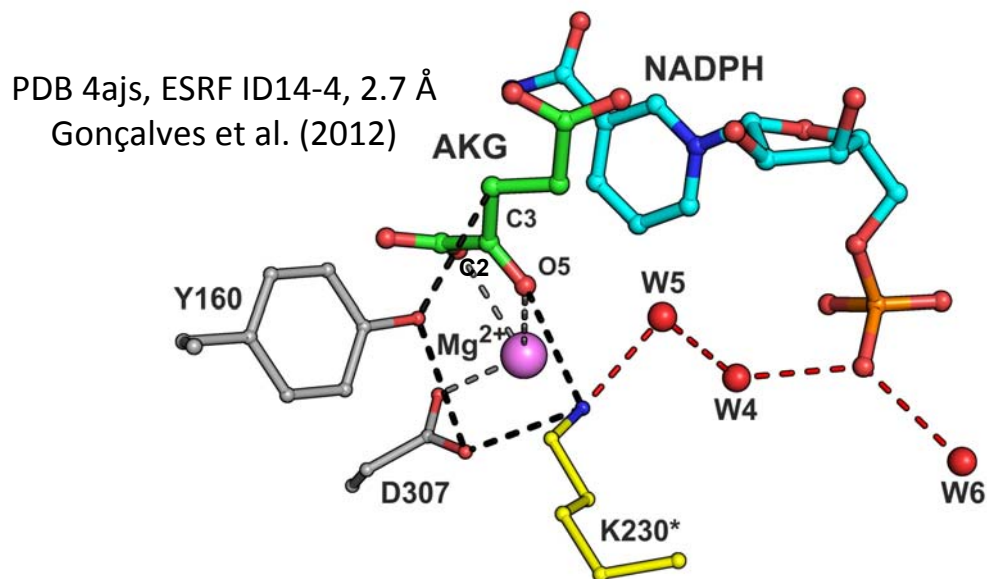
Changes in the electrostatic potential landscape of the *E. coli wt* IDH active site from the **open** (left) to the **quasi-closed** (centre) and **fully-closed** (right) conformations.

The motion of the “NADP loop” and “P loop” is also evident.



Induced Fit and the Catalytic Mechanism of Isocitrate Dehydrogenase

The product complex **K100M IDH:NADPH:AKG:Ca²⁺** was obtained by ICT turnover *in crystallum*

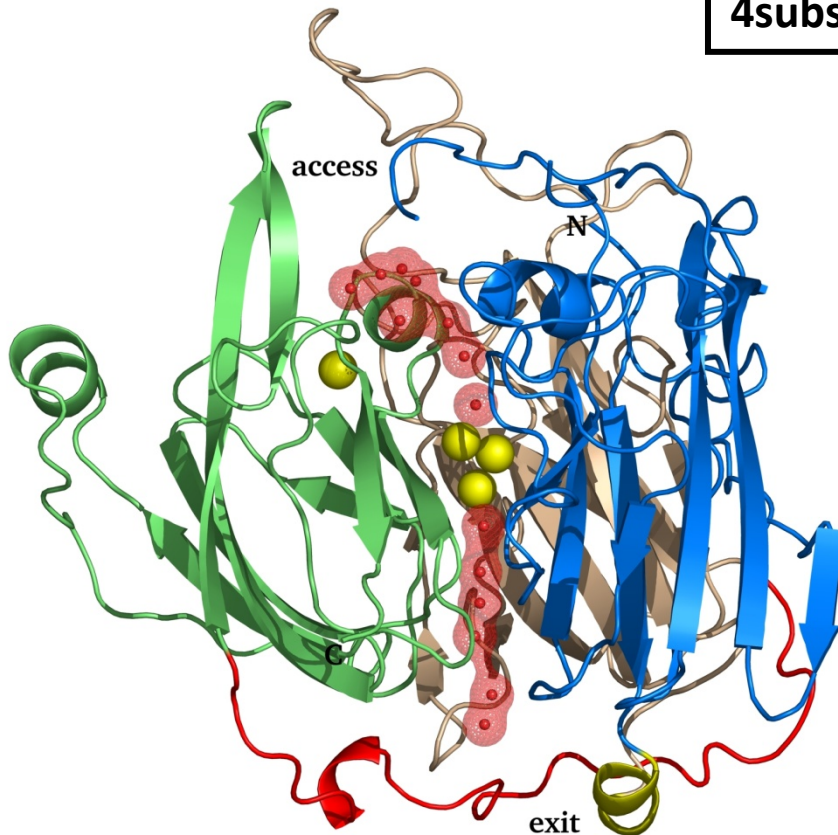
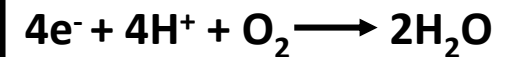
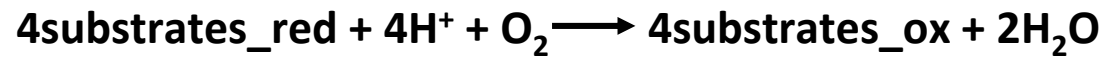


Y160 after C3 protonation (d = 3.4 Å)

K230* after C2 hydroxyl deprotonation (O5 in AKG) (d = 3.3 Å)

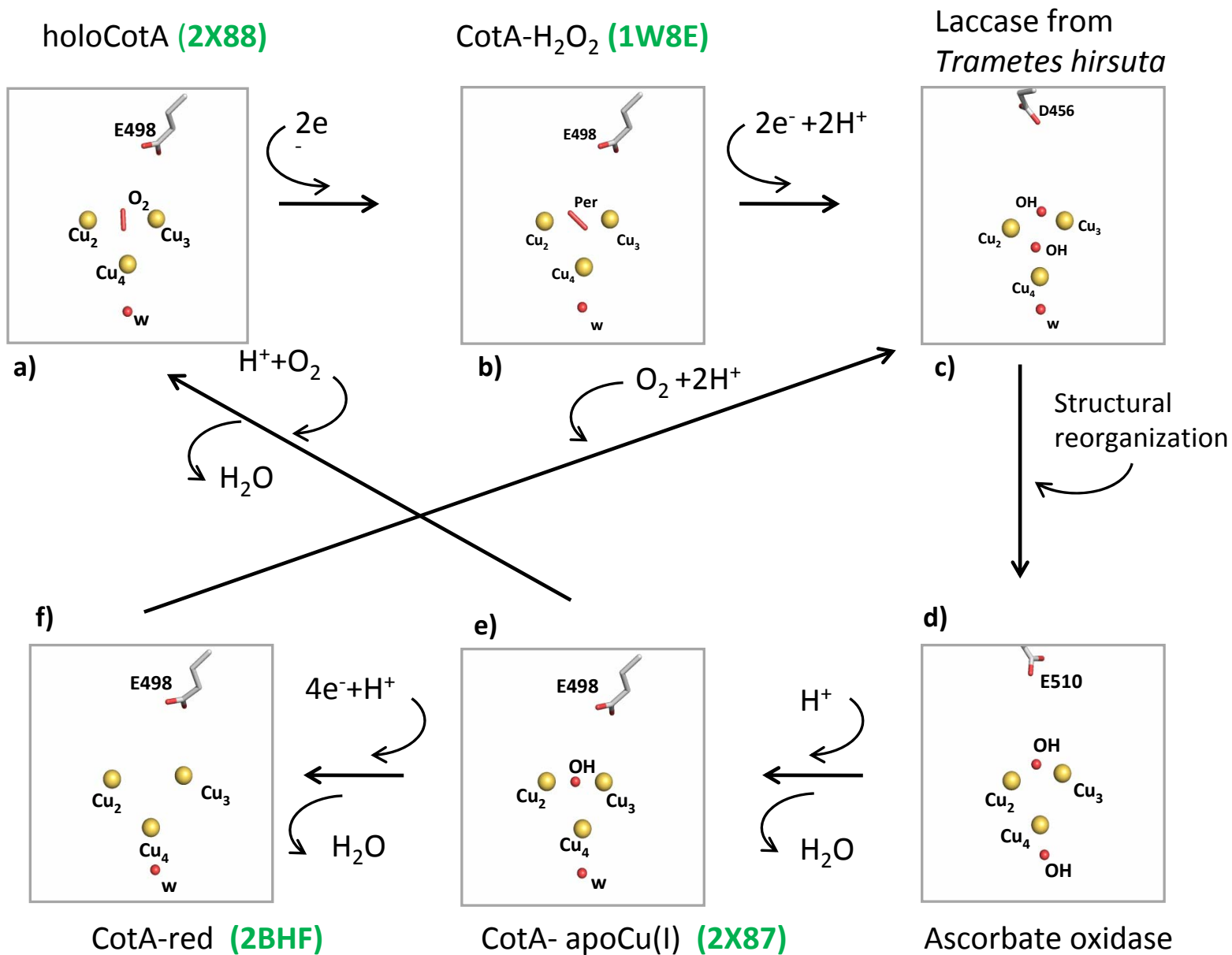
Y160 re-protonation: D307 or a proton relay from bulk solvent (I, II)

Dioxygen reduction by Multicopper oxidases



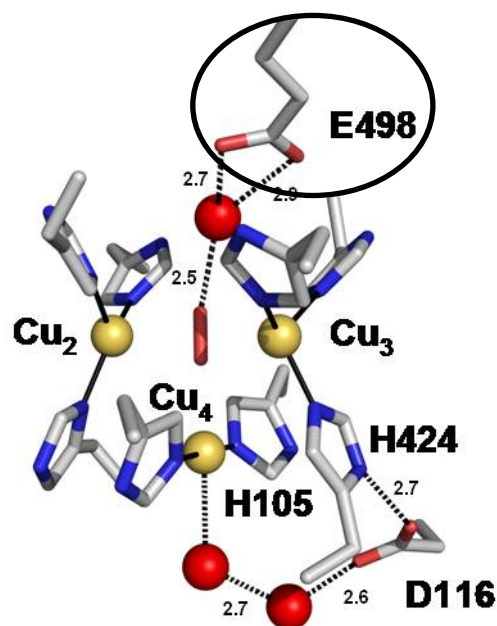
- Which oxygen species are present in the mechanism?
- Which residues are involved in the proton transfer events that occur during the mechanism?

Proposed mechanism for dioxygen reduction by multicopper oxidases

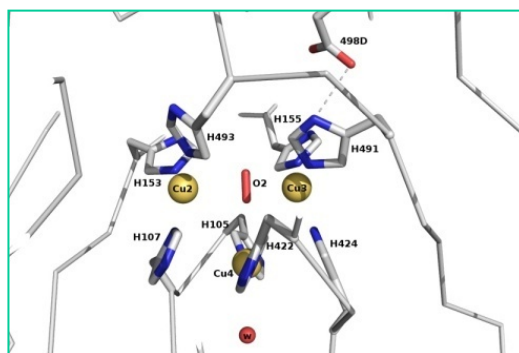


Which residues are involved in proton transfer?

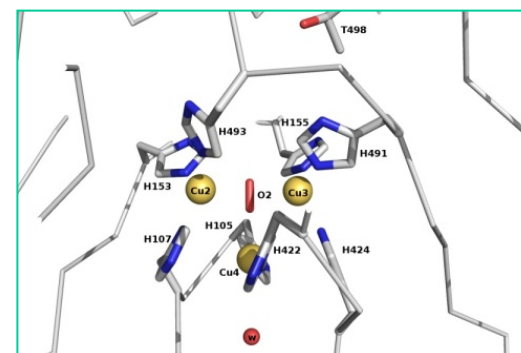
Site-directed mutagenesis of E498



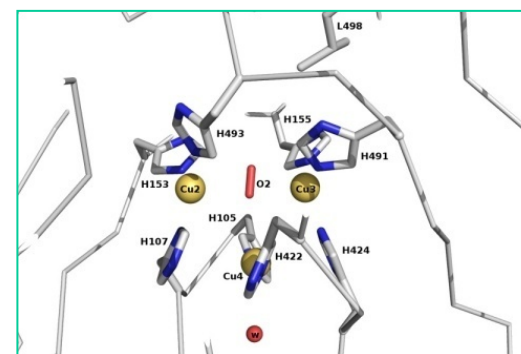
E498D (4AKQ)



E498T (4AKP)



E498L (4AKO)

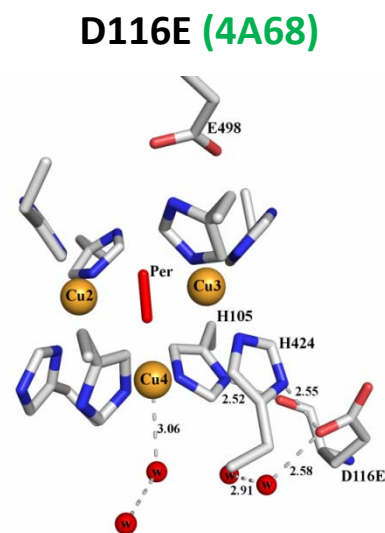
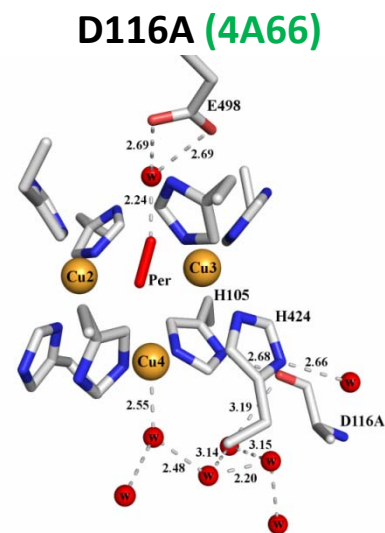
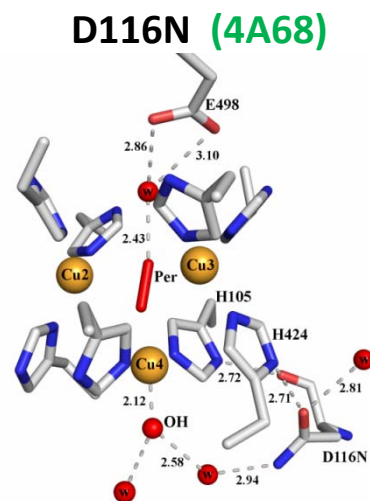
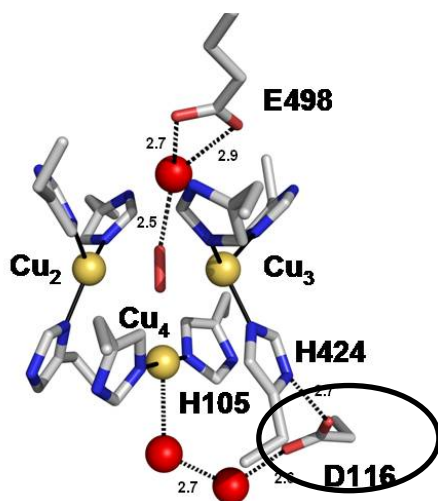


Important differences were located at the copper centres and in their neighbourhood.

498T and 498L mutants were inactive and 498D retained part of the catalytic activity showing that Glu498 plays an important role in channelling the protons to the mechanisms of dioxygen reduction and that no other alternative pathway is observed.

Which residues are involved in proton transfer?

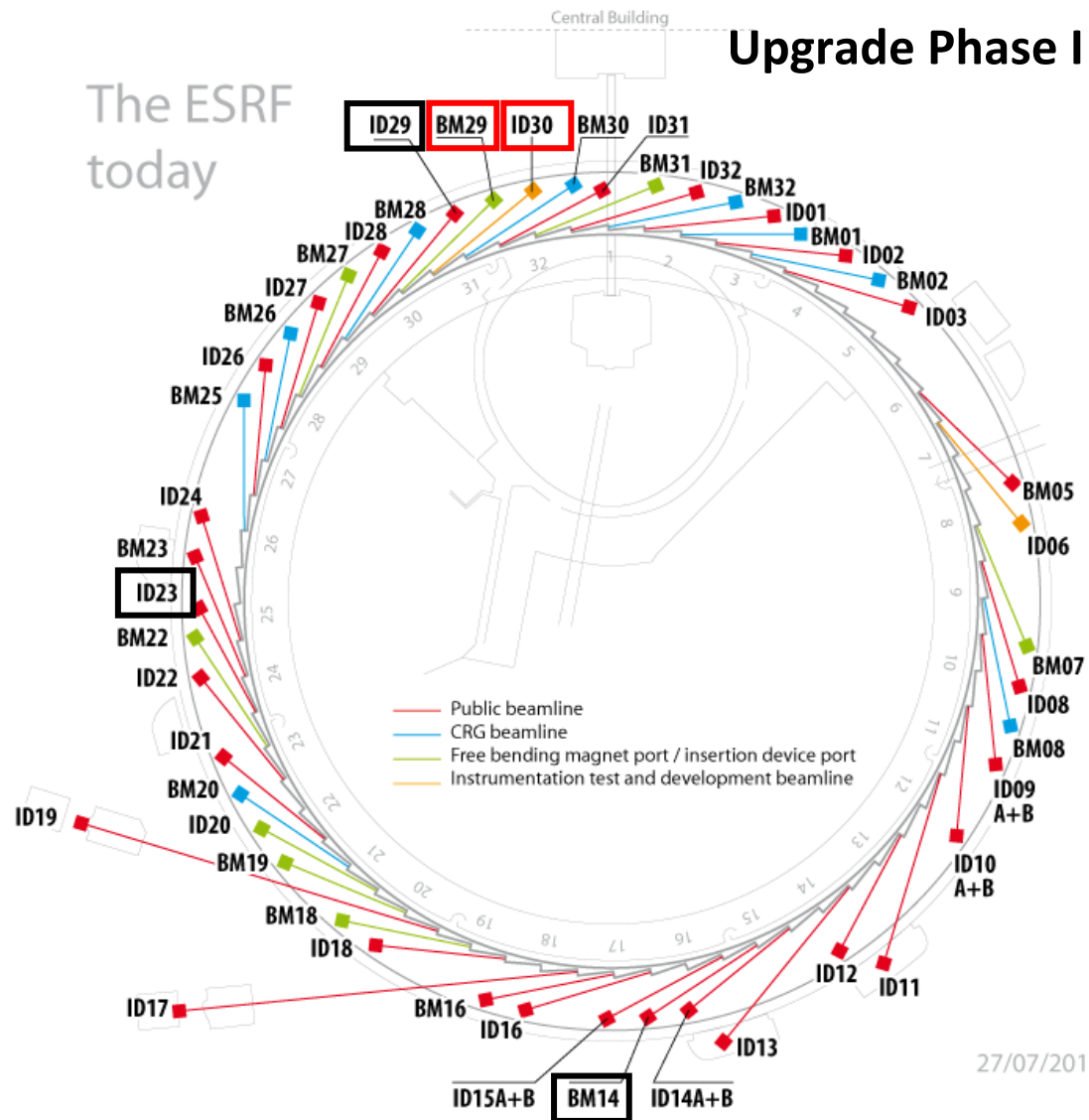
Saturated mutagenesis of D116



The structures of the mutant enzymes D116N, D116A and D116E, were used in equilibrium protonation simulations in order to further assess the role of Asp116 during the protonation process.

The results indicate that D116 is important for catalysis, either by modulating the protonation events through E498 or by maintaining the local geometry and water connectivity at the trinuclear copper site.

Things to come at the ESRF



Things to come at the ESRF

Upgrade Phase II:

- *Architecture of the straight sections will be changed to reduce beam horizontal emittance → Long (1-year) shutdown in 2018-2019*
- *Increase in X-ray brilliance/flux:*
 - *10-fold at the BM beamlines*
 - *Up to 40-fold at the ID (Undulator) beamlines*
- *Radiation damage problems increased → crystal lifetime may be as low as a few milliseconds!*
- *New methods for data collection and sample handling are in development*

Things to come at the ESRF



User input and participation at the Users Meeting and Associated Workshops is very important!